

## Assessment of hepatoprotective activity of *Caralluma cicatricosa* against CCl<sub>4</sub>-induced liver damage in rabbits

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### ABSTRACT

Liver disease has become one of the serious health problems as it is exposed to many kinds of xenobiotics and therapeutic agents. Certain medicinal plants had been used to cure some liver diseases. The present study aimed to evaluate the hepatoprotective effect of methanolic extract of *Caralluma cicatricosa* (MECC) in rabbits with acute liver injury induced by carbon tetrachloride at a single dose of 1.25 ml/kg b.w. as a mixture with olive oil. MECC was administered in doses of (0.5, 1 and 1.5 g/kg b.w.) *via* oral gavage by intragastric tube for 8 days. The SGOT, SGPT, ALP, total protein, creatinine, blood urea nitrogen and blood glucose level were measured and histopathology of liver was performed. The results of the rabbits treated with MECC were compared with that of Liv-52. *C. cicatricosa* showed a significant dose-dependent reduction ( $P < 0.05$ ) in the hepatic enzymes levels, blood urea nitrogen, blood glucose level and improvement of serum protein. The histopathological studies in the liver of rabbits also supported that *C. cicatricosa* extract markedly reduced the toxicity of CCl<sub>4</sub> and preserved the histoarchitecture of the liver tissue to near-normal structure. These results suggested that *C. cicatricosa* may be acting as a natural hepatoprotective agent that prevents hepatic injury induced by CCl<sub>4</sub> and this may be due to its active phytoconstituents such as flavonoids and glycosides which present in the plant.

**Keywords:** Carbon tetrachloride, *Caralluma cicatricosa*, Hepatoprotective, Rabbits.

### INTRODUCTION:

The liver is one of the important organs in the body. It plays a pivotal role in regulating various physiological processes. It helps in the maintenance, performance and regulating homeostasis of the body. It is involved in almost all the biochemical pathways to growth, fighting against disease, nutrient supply, energy provision and reproduction. In addition, it aids metabolism of carbohydrate, protein and fat detoxification, secretion of bile and storage of vitamins [1]. Unfortunately, the liver is often abused by environment, toxins, alcohol and over-the-counter drug use (xenobiotics) which can damage the liver and eventually lead to hepatitis, cirrhosis and liver diseases [2]. About 25,000 deaths found every year due to liver disorders [3]. Therefore, treatment of liver diseases is extremely important. Medicinal plants have been used for the treatment of several human diseases over the centuries and have been very important in the health care delivery of every nation [4].

The genus *Caralluma* comprises about 200 species distributed throughout Africa and Asia. The majority of

these species are indigenous to the Indian sub-continent and Arabian Peninsula [5]. The etymology of the word "*Caralluma*" is from the Arabian, "qarh al-luhum", meaning wound in the flesh or abscess, probably from the floral odor in some taxa [6]. Some species of *Caralluma* are edible and form part of the folk medicine system of many countries [7]. These are commonly used in traditional medicine as remedies to relieve wide variety of illness and health problems [8]. *Caralluma* species have shown anti-inflammatory, anti-nociceptive [9,10], antidiabetics [11,12], hepatoprotective [13], gastric mucosa protecting [14], antimalarial [15], anti-oxidant [16], anti-trypanosomal [15,17], appetite suppressant [18] and cytotoxic [19,20,21,22] activities.

*Caralluma cicatricosa* is belonging to the family of Asclepiadaceae, which is also known as the milkweed family because many of its members contain milky latex [23]. *Caralluma cicatricosa* is a succulent and angular plant which growing wild in western and Arabian Peninsula. In Yemen it's found in Taiz, Maaber, Milhan, Manakha and mountain Arays [24]. In traditional Yemeni folk medicine

*C. cicatricosa* is used in case of diabetes and in peptic ulcer and its juice as drops for ear inflammation. There are no phytochemical and pharmacological work has been done on *C. cicatricosa*. In view of traditional uses as well as the diverse biological activities so far reported for different *Caralluma* species, the *C. cicatricosa* was chosen.

CCl<sub>4</sub> is a well-known hepato- and nephrotoxicant [25,26,27,28] and proves highly useful as an experimental model for the study of certain hepatotoxic effects [29,30]. The changes associated with CCl<sub>4</sub>-induced liver damage are similar to that of acute viral hepatitis [31].

## MATERIALS AND METHODS

### Drugs and Chemicals

Drugs and reagents were used include: Carbon Tetrachloride from (BDH Chemicals Ltd, Poole, England), standard Kits for assay of Aspartate transaminase (AST), Alanine transaminase (ALT), Alkaline phosphatase (ALP) activities in addition to, total protein, urea and creatinine levels were purchased from (SPINREACT, Spain). Liv-52 (Himalaya Drugs, India) from local supplier. All other reagents and solvents were of highest purity and analytical grade.

### Plant Collection and identification

The Fresh plant was collected from various locations around Ibb city in April and May 2014 and authenticated by the taxonomist Prof. Abdulnaser Al Gifri from the department of biology, faculty of education, Aden University, Yemen.

### Plant extraction

The plant roots were removed and stem (bark) of plant were washed in floating water, cut into smaller pieces and dried under shade for 14 consecutive days. The dried stems of plant were grounded with a mechanical mill (SM MARSHAL-SUPERY DRY GRINDER-MODEL NO:SM-830) and passed through sieves (No-60), to obtain fine mass. The powdered material (256.8 g) was cold macerated in 70% v/v Methanol in conical flask (V1000 ml) for three days (72 hours) with frequent stirring. Then the extract was filtered using cotton and concentrated under vacuum at 40°C using a rotary evaporator (Buchi-type) and kept in closed container at approximately -20 °C.

### Plant extracts yield and phytochemical analysis

The yield percentage of the extract was calculated using the following equation [32].

$$\text{The Yield (\%)} = \frac{\text{Weight of the extract (g)}}{\text{Weight of the ground plant material (g)}} \times 100$$

The 70% methanolic bark extract was subjected to a preliminary phytochemical screening for the presence of tannins, alkaloids, flavonoids, glycosides (reducing sugars), anthraquinones, sterols and saponins by simple qualitative methods [33,34].

### Experimental animals

Adult healthy rabbits of weights (900g-1200g) and aged 8 month ± 1 week were obtained from a local supplier, city of IBB (IBB, Yemen). Animals were housed in metal cages (80×80×50 cm) with soft wood shavings as bedding and fed with clean water and food throughout the period of the experiment. The animals were acclimatized for one week before starting the experiment. The study protocol was approved by the University Ethics committee before any experiment can be conducted.

### Acute toxicity study

Acute toxicity study was conducted by 5 female rabbits according to the Organization for Economic Cooperation and Development (OECD) revised up-and-down procedure for acute toxicity testing (OECD guideline 425, 2001) [35]. Non-pregnant female rabbits weighing 900-1000g were divided and housed singly in metal cages for one week before the start of experiment. Rabbits were fasted overnight from food, but not water, prior to dosing and weighed before the extract was administered orally. A dose of 5000 mg/kg was dissolved in distilled water at an administration volume of 10 ml/kg body weight based on the individual animal body weights obtained on the day dosing and given to the first rabbit by oral gavages, and this rabbit was observed for mortality and clinical signs for the first hour, then hourly for 3h and finally periodically until 48h. If the animal survived, then four additional animals were given the same 5000 mg/kg dose sequentially at 48-h intervals.

All of the experimental animals were maintained under close observation for 14 days, and the number of rabbits that died within the study period was noted. The LD<sub>50</sub> was predicted to be above 5000 mg/kg if three or more rabbits survived.

### Experimental design

Forty two male rabbits were grouped into seven groups each contain six animals. Group (I) served as the normal control and received only 0.5% CMC (carboxymethyl-cellulose) suspension by a gastric gavage throughout the experimental period. Group (II) treated by methanolic extract of *C. cicatricosa* (MECC) 1000 mg/kg b.w. Group (III) served as the toxin control and received a 0.5% CMC suspension by a gastric gavage. Group (IV), (V), and (VI) received MECC 500 mg/kg b.w., 1000 mg/kg b.w. and

1500 mg/kg b.w. respectively while animals in group (VII) received liv-52 (5ml/kg b.w.) [39] for seven consecutive days. On the eighth day, animals groups III, IV, V, VI & VII were given single oral dose of CCl<sub>4</sub> (1.25 ml/kg) [36,37,38] diluted in olive oil at rate of 1:1(v:v) Sixty minutes after the last administration of the vehicle, extract and liv-52 [40].

### Blood Collection

Twenty four hours later [41,42], overnight fasting rabbits were deprived of food but allowed for free access of drinking water. Animals were anaesthetized with ether and blood was collected by direct cardiac puncture into tubes without anticoagulant and allowed for 30 minutes to clot at room temperature. After the clotting of the blood, it was centrifuged at 4000 rpm for ten minutes (HettichMikro 220R centrifuge, DJB lab care, UK) at 25°C for separation of serum. The supernatant was stored in the freezer at -21°C until analysis [43].

### Biochemical analysis

Serum levels of alanine transaminase (ALT), aspartate transaminase (AST), alkaline phosphatase (ALP), total proteins, creatinine, urea and glucose were determined kinetically using commercially available kits.

### Histopathological Examination

At the end of the experimental period (8 days), rabbits were anesthetized and sacrificed; liver was removed and kept in 10% formaldehyde solution and send to University of Science and Technology hospital laboratory were tissue processed and stained with hematoxyline and eosin for histological examination [44,45].

### Statistical analysis

Statistical analysis of the results was done using the computer software program "SPSS v.21". Comparison between different groups was carried out using the one-way analysis of variance (ANOVA) followed by least significant difference test (LSD test). Difference was considered significant when P is less than 0.05.

## RESULTS

### Plant extracts yield and preliminary phytochemical analysis

The percentage yield of the 70% methanolic crude extract which was obtained by cold extraction was (21.18% w/w). The results of the phytochemical screening revealed the presence of steroids, flavonoids, tannins, glycosides, saponins, carbohydrates and triterpenes in the 70% of MECC steam. Alkaloids and anthraquinones were not found in the tested extract.

### Acute toxicity

The single oral administration of MECC in rabbits was non-toxic up to a dose of (5000 mg/kg) as no mortality was detected at the end of experiment (14 days).

### Biochemical tests

The results showed that CCl<sub>4</sub> produced a significant increase in AST, ALT & ALP levels and a significant reduction in the total protein level, indicating hepatocellular damage. On other hand, administration of MECC at the doses of 500mg, 1000mg and 1500 mg/kg b.w. produced a highly significant fall in the AST, ALT & ALP levels and a significant rise in the total protein levels in a dose dependent manner. Similar results were also obtained with liv-52+CCl<sub>4</sub> which considered as a standard group. There was no significant alteration in control rabbits treated solely with MECC. (Table 1)

The administration of CCl<sub>4</sub> alone induced a significant increase in blood urea nitrogen (BUN), creatinine levels compared to control group. on other side, orally pretreatment of animals with MECC+CCl<sub>4</sub> and Liv-52+CCl<sub>4</sub> reduced the level of blood urea nitrogen significantly in these groups with respect to rabbits intoxicated with CCl<sub>4</sub>. However, there is no significant decrease in creatinine level in animals pretreatment with MECC+CCl<sub>4</sub> and MECC. The result showed significant increase in blood sugar level in animal models treated with CCl<sub>4</sub> compared with control (Table 2). Similarly, there were significant reductions in blood sugar values of animals pre-treated with MECC+CCl<sub>4</sub> and Liv-52+CCl<sub>4</sub> compared with CCl<sub>4</sub> treated group. Additionally, the level of glucose in the animals that pretreated with MECC, MECC+CCl<sub>4</sub> and Liv-52 was significantly decreased compared to normal group.

### Histopathology findings

Liver morphological changes were scored, described and summarized in control, MECC treated, CCl<sub>4</sub> treated, CCl<sub>4</sub>+MECC treated & Liv-52 treated groups (Table 3). Liver sections of control group and rabbits pretreated with MECC showed normal cellular architecture with distinct hepatic cells, sinusoidal spaces, and central vein. However, CCl<sub>4</sub> application constituted histopathological changes in the liver of rabbits. These changes were characterized by dilation of central vein with noticeable hemorrhage. Moreover, hepatocyte necrosis, inflammatory cells infiltration, fatty degeneration, and hydropic degeneration were found in rabbits 24 hours after CCl<sub>4</sub> treatment (Figure 1). However, liver sections of the rabbits received MECC and Liv-52 prior to CCl<sub>4</sub> intoxication showed less histopathological changes in a dose dependent manner when compared to CCl<sub>4</sub> group.

**Table1: The effect of methanolic extract of *Caralluma cicatricose* associated with CCl<sub>4</sub>-induced hepatic damage on liver function tests in rabbits.**

Group	ASTU/L	ALTU/L	ALPU/L	TP g/dL
Normal control (5%CMC5mL/kg p.o.)	80.250±1.956	71.100±6.049	130.250±6.503	5.450±0.085
MECC (1g/kg p.o.)	75.250±0.540	57.350±4.148	118.500±5.529	5.250±0.140
CCl <sub>4</sub> (1.25ml/kg p.o.)	395.667±31.110*	438.817 ±43.331*	441.333±32.368*	4.083±0.429*
CCl <sub>4</sub> +MECC (0.5g/kg p.o.)	284.267±54.930* <sup>#</sup>	341.500 ±21.603* <sup>#</sup>	225.767±18.003* <sup>#</sup>	5.150±0.232 <sup>#</sup>
CCl <sub>4</sub> +MECC (1g/kg p.o.)	264.500±37.825* <sup>#</sup>	290.800 ±41.201* <sup>#</sup>	223.800±29.091* <sup>#</sup>	5.250±0.096 <sup>#</sup>
CCl <sub>4</sub> +MECC (1.5g/kg p.o.)	225.667±17.091* <sup>#</sup>	235.267 ±39.457* <sup>#</sup>	181.000±24.324* <sup>#</sup>	5.317±0.168 <sup>#</sup>
CCl <sub>4</sub> +liv52 (5ml/kg p.o.)	190.350 ±12.766* <sup>#</sup>	175.880 ±24.279* <sup>#</sup>	205.800±26.164* <sup>#</sup>	5.483±0.140 <sup>#</sup>

Data expressed in mean±SEM, n=6. \*: P<0.05, significant difference from the normal control group; #: P<0.05, significant difference from the CCl<sub>4</sub> group

**Table 2: The effect of methanolic extract of *Caralluma cicatricose* associated with CCl<sub>4</sub> toxicity on kidney function tests and Fasting blood sugar in rabbits.**

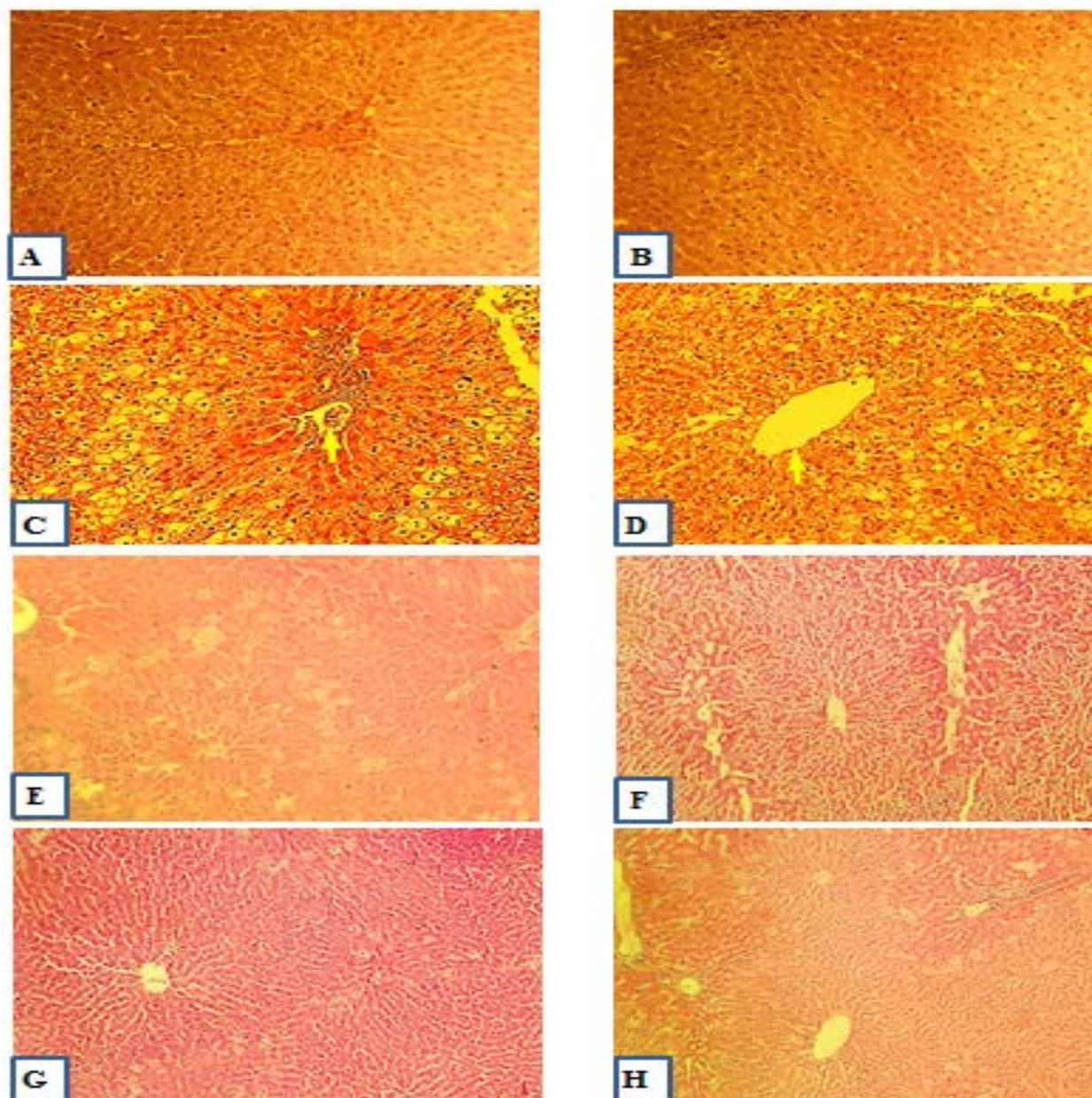
Group	Blood urea nitrogen(mg/dL)	Creatinine (mg/dL)	Fasting blood sugar (mg/dL)
Normal control (5% CMC 5mL/kg p.o.)	55.450±3.593	0.958±0.013	137.750±5.522
MECC (1g/kg p.o.)	58.583±4.669	1.023±0.019	90.250±9.975*
CCl <sub>4</sub> (1.25ml/kg p.o.)	75.667±7.805*	1.417±0.185*	153.000±15.438*
CCl <sub>4</sub> +MECC(0.5g/kgp.o.)	49.000±1.414 <sup>#</sup>	1.110±0.066	89.833±1.537* <sup>#</sup>
CCl <sub>4</sub> +MECC(1g/kg p.o.)	50.667±3.018 <sup>#</sup>	1.260±0.117	88.167±1.447* <sup>#</sup>
CCl <sub>4</sub> +MECC(1.5g/kgp.o.)	53.167±2.960 <sup>#</sup>	1.400±0.328*	83.333±3.383* <sup>#</sup>
CCl <sub>4</sub> +liv52 (5ml/kg p.o.)	47.167±0.703 <sup>#</sup>	0.948±0.057 <sup>#</sup>	86.517±2.696* <sup>#</sup>

Data expressed in mean±SEM, n=6. \*: P<0.05, significant difference from the normal control group; #: P<0.05, significant difference from the CCl<sub>4</sub> group

**Table 3: Histological injury scores of liver under different doses of MECC in rabbits treated with CCl<sub>4</sub>.**

Group	Injury Score <sup>a</sup>				
	Fatty degeneration	Architecture distortion	congestion	Necrosis	Inflammation
Control	0	0	0	0	0
MECC (1 g/kg b.w.)	0	0	0	0	0
CCl <sub>4</sub> (1.25ml/kg)	4	3	3	3	3
MECC (0.5 g/kg + CCl <sub>4</sub> )	3	3	2	2	2
MECC (1 g/kg + CCl <sub>4</sub> )	2	2	2	2	1
MECC (1.5 g/kg + CCl <sub>4</sub> )	1	1	2	3	3
Liv-52 (5mL/kg + CCl <sub>4</sub> )	1	1	2	2	2

<sup>a</sup> Livers were scored for hepatic injury via light microscopy with score 0 = no visible cell damage; score 1 = focal hepatocyte damage on less than 25 % of the tissue; score 2 = focal hepatocyte damage on 25-50 % of the tissue; score 3 = extensive but focal with hepatocyte lesions; score 4 = global hepatocyte necrosis.



**Figure 11:** Photographs of liver sections of  $\text{CCl}_4$  (1.25 ml/kg) toxicity in rabbits (hematoxylin and eosin stained, 10x). **(A)** Normal control, **(B)** MECC 1 gram/kg, **(C&D)** Toxin control ( $\text{CCl}_4$ ), **(E)** MECC 0.5 gram/kg +  $\text{CCl}_4$ , **(F)** MECC 1 gram/kg +  $\text{CCl}_4$ , **(G)** MECC 1.5 gram/kg and **(H)** Liv-52 5ml/kg +  $\text{CCl}_4$ .

### Discussion

Phytochemical screening of *C. cicatricosa* has revealed the presence of steroids, flavonoids, tannins, glycosides, saponins, carbohydrates and triterpenes. Phytoconstituents like flavonoids, triterpenoids, saponins and alkaloids are known to possess hepatoprotective activity [46,47]. *C. cicatricosa* possesses a significant protective effect against hepatotoxicity induced by  $\text{CCl}_4$  which may be attributed to the individual or combined action of phytoconstituents present in it. The liver-

protective herbal drugs have been shown to contain a variety of chemical constituents like phenols, coumarins, ligands, essential oils, monoterpenes, carotenoids, glycosides, organic acids, lipids, xanthene etc. [48,1]. Thus it is possible that other secondary metabolites of the plant as observed in the present study may be responsible for the hepatoprotective activity of the plant. In the current study, acute toxicity test was done to establish if any adverse effects of the administration of MECC on some observable parameters. Changes in body

weight have been used as an indicator of adverse effects of drugs and chemicals[49,50]. No significant changes were observed in the general behavior, body weight, suggesting that at single oral doses administered, MECC had no effect on the normal growth of rabbits. From the results of acute oral toxicity study of the methanolic extract, it can be concluded that LD50 of the drug is greater than 5000 mg/kg bodyweight, which means that even a dose of 5000 mg/kg bodyweight of methanolic extract of stem of *C. cicatricosa* is safe for administration. CCl<sub>4</sub>-induced hepatotoxicity is believed to involve two phases. The initial phase involves the metabolism of CCl<sub>4</sub> by cytochrome P450 to the trichloromethyl radicals (CCl<sub>3</sub><sup>•</sup> and/or CCl<sub>3</sub>OO<sup>•</sup>)[51,52], a free radical that binds to lipoprotein and leads to peroxidation of lipids of the endoplasmic reticulum which causes changes in the physical and chemical properties of cellular membranes[53,54], thus effecting their fluidity and severity of liver injury[63]. The elevated activities of these enzymes are indicative of cellular leakage and loss of the functional integrity of the cell membranes in liver which is always associated with hepatonecrosis[64,65]. In this study, significant increase in AST, ALT and ALP levels in the serum was observed after oral administration of CCl<sub>4</sub>, indicated that CCl<sub>4</sub> intoxication compromised the integrity of the hepatic cell membranes [66,67,68]. Pretreatment with MECC+CCl<sub>4</sub> and Liv52+CCl<sub>4</sub> significantly decreased levels of AST, ALT and ALP towards the normal value. These observations are in agreement with the commonly accepted view that serum levels of transaminases return to normal due to stabilization of plasma membrane as well as repair of hepatic tissue damages caused by CCl<sub>4</sub>[69].

Reduction in total protein content can be deemed as a useful index of severity of hepatocellular damage[70]. In the present study, CCl<sub>4</sub> intoxication reduced the serum total protein level which is attributed to the initial damage produced and localized in the endoplasmic reticulum which results in the loss of cytochrome P-450 enzymes leading to its functional failure with a decrease in protein synthesis and accumulation of triglycerides leading to fatty liver[71]. Previous studies have showed that administration of CCl<sub>4</sub> to a diversity of animal types results in a rapid reduction in protein synthesis in the liver [66,72,73]. The pretreatment of MECC+CCl<sub>4</sub> restored the total protein level that suggests the stabilization of endoplasmic reticulum leading to protein synthesis[59].

The kidney assistances in keeping homeostasis of the body by reabsorbing important material and evacuating

permeability for ion exchange, resulting in leakage of enzymes in blood and finally results in swelling, cytolysis, and cell death[55,56,57]. The second phase of CCl<sub>4</sub>-induced hepatotoxicity involves the activation of Kupffer cells, which is accompanied by the production of proinflammatory mediators[58]. The effect of CCl<sub>4</sub> is generally observed after 24 h of its administration and then values slowly normalized. Hence the withdrawal of the blood for biochemical parameters should be carried out only after 24 h of CCl<sub>4</sub> intoxication[59,42].

The liver marker enzymes (AST, ALT and ALP) are largely used as most common biochemical markers to evaluate liver injury[60,61] because these enzymes are placed in cytoplasmic area of the cell and are released into circulation in case of cellular damage[62]. Thus, the activities of these enzymes in serum can reflect the

waste yields. Creatinine is a commonly used as measure of kidney function[74]. The increase in the level of creatinine in the blood considered an indicator to the kidney damage[75]. In this experiment, oral administration of CCl<sub>4</sub> resulted in a significant increase in creatinine and blood urea nitrogen concentrations. The observed increase is indicative of cellular leakage and loss of functional integrity of cell membrane in renal tissue[76,77]. In the current study, the pretreatment with MECC+CCl<sub>4</sub> significantly decreased serum urea levels in a dose dependently manner, compared to CCl<sub>4</sub>-induced rabbits by enhancing the renal function that is generally impaired in CCl<sub>4</sub>-induced rabbits. On other hand, elevated serum level of creatinine in the CCl<sub>4</sub> group in this study was found to reduce insignificantly sequel to administration of MECC in a dose-dependent manner. But the dosage of 1500mg/kg b.w. increased serum creatinine level significantly compared to normal model. This suggests that the high dose of this plant extract may precipitate kidney damage.

In the present study, serum glucose level was higher in CCl<sub>4</sub> treated rabbits. The elevation of glucose level could be recognized to destruction of hepatocytes brought by CCl<sub>4</sub> intoxication[78,79] or decreasing of glycogen contents due to degradation of glycogen to glucose in hepatocytes after treatment with CCl<sub>4</sub> which leads to increase in glucose level in the blood[80]. The hypoglycemic effect of *C. cicatricosa* observed in the present work was consistent with the observation on other *Caralluma* extract[11]. The possible mechanism may be by potentiation of insulin effect either by increasing the pancreatic secretion of insulin from β-cells

of islets of Langerhans or by increasing the peripheral glucose uptake[81].

Liv-52 is widely used in treatment of liver diseases of varying origins. It enhances tocopherol levels, which inhibits lipid peroxidation and scavenges free radicals[82]. In the present study, Liv-52 caused a significant decrease in serum enzymes activity induced by CCl<sub>4</sub> in rabbits. These results were in agreement with previous investigation[39].

On the contrary, pre-treatment with MECC alone did not show any significant elevation in the amino transferases activities and kidney biochemical parameters when compared to control. This finding is another pointer to the safety of this extract for consumption at this concentration.

Histopathological study showed extensive fatty degeneration, architecture distortion, congestion, necrosis and inflammation which successfully induced by oral administration of single dose of CCl<sub>4</sub>[83,84]. The histological appearance of MECC and liv-52 groups was partly similar to that of the control group, and tissue damage and necrosis were of less extent in these groups than the CCl<sub>4</sub> group in a dose-dependent manner. As well as, no abnormalities or no changes observed in liver sections of rabbits pretreated by MECC and this indicated safety of the plant on liver organ. The overall histopathological findings are well correlating with the biochemical estimations and suggested that *C. cicatricosa* may be effective against CCl<sub>4</sub>-induced changes in liver.

## CONCLUSIONS

This study has shown that the methanolic extract of *Caralluma cicatricosais* is safe in experimental rabbits up to a dose of 5000 mg/kg and has hepatoprotective activity against carbon tetrachloride-induced liver damage. This activity of the extract may be due to the presence of flavonoids and other phytochemical constituents present in the plant.

## FUTURE WORK

Recommended future work is to look at the effect of *C. cicatricosa* on diabetic animals as well as to isolate the chemical compound(s) responsible for hepatoprotection. Moreover, sub-acute and chronic evaluation of the extract should be carried out in evaluating the safety profile of *C. cicatricosa*.

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## CONFLICT OF INTERESTS

There is no conflict of interests regarding the publication of this paper.

## REFERENCES

1. Ahsan MR, Islam KM, Bulbul IJ. Hepatoprotective activity of methanol extract of some medicinal plants against carbon tetrachloride-induced hepatotoxicity in rats. *Eur J Sci Res* .2009; 37(2): 302-310.
2. Siripong P, Kongkathip B, Preechanukool K, Pich P, Tunsuwan K, Taylor WC. Cytotoxic diterpenoid constituents from *Androgra phispaniculata* Nees leaves. *J. Sci.Soc. Thailand*, .1992;18(4):187-94.
3. Sharma B, Sharma UK. Hepatoprotective activity of some indigenous plants, *Int J Pharma. Tech. Res*. 2010;2:568-572..
4. Oluma HO, Umoh EU, Onekutu A, Okolo J. Antibacterial potentials of eight medicinal plants from the lower Benue valley of Nigeria against *Salmonella typhi*., *Niger. J. Bot*. 2004;17: 1-11.
5. Gilbert MG.A review of *Caralluma R. Br.* and its segregates, *Bradleya*.1990;8: 1-32
6. Albers F, Meve U. Illustrated handbook of succulent plants: Corrected 2nd printing-Verlag, Springer. Berlin, Heidelberg, New York;2004.
7. Abdel-Sattar E, Ahmed AA, Hegazy ME, Farag MA, Al-Yahya MA. Acylated pregnane glycosides from *Caralluma russeliana*. *Phytochemistry*.2007;68:1459-1463.
8. Ahmad MM, Qureshi S, Shah A, Qazi NS, Rao RM, Al-Bekairi AM. Anti-Inflammatory activity of *Caralluma tuberculata* alcoholic extract. *Fitoterapia*. 1983;46: 357-360.
9. Rammesh M, Roa YN, Roa AV, Prabhakar MC, Roa SC, Muralidhar N, Reddy BM. Antinociceptive and anti-inflammatory activity of a flavonoid isolated from *Caralluma attenuate*. *J Ethnopharmacol*. 1998;62:63-66.
10. Zakaria MN, Isran MW, Radhakrishnan R. et al. Antinociceptive and anti-inflammatory activity properties of *Caralluma arebica*. *J Ethnopharmacol*. 2001;76:155-158.
11. Habibuddin M, Dagherri HA, Humaira T, Al Qahtani MS, Hefzi AA. Antidiabetic effect of alcoholic extract of *Caralluma sinaica*. on streptozotocin induced diabetic rabbits. *J Ethnopharmacol*.2008;117:215-220.

12. Venkatesh S, Reddy GD, Reddy BM, Ramesh M, Roa AV. Antihyperglycemic activity of *Caralluma attenuate*. *Fitoterapia*.2003;74(3):274-279.
13. Shanmugam G, Ayyavu M, Rao DM, Devarajan T, Subramaniam G. Hepatoprotective effect of *Caralluma umbellate* against acetaminophen induced oxidative stress and liver damage in rat .J Pharm Res.2013;6(3):342–345.
14. Al-Harbi MD, Qureshi S, Ahmed MM, Afzal M, Shah AH. Evaluation of *Caralluma tuberculata* pretreatment for the protection of rat gastric mucosa against toxic damage.*ToxicolAppl Pharmacol*.1994; 128:1-8.
15. Abdel-Sattar E, Harraz FM, Al-Ansari SM, El-Mekkawy S, Ichino C, Kiyohara H, Ootoguro K, Omura S, Yamada H. Antiplasmodial and antitrypanosomal activity of plants from the Kingdom of Saudi Arabia.*J Nat Medi*.2009;63: 232-239
16. Ansari NM, Houlihan L, Hussain B, Pieroni A. Antioxidant activity of five vegetables traditionally consumed by South-Asian migrants in Bradford, Yorkshire, UK.*Phytother. Res*.2005;19:907-911.
17. Abdel-Sattar E, Shehab NG, Ichino C, Kiyohara H, Ishiyama A, Ootoguro K, Omura S, Yamada H. Antitrypanosomal activity of some pregnane glycosides isolated from *Caralluma* species.*Phytomedicine*.2009;16:659-664.
18. Kunert O, Rao BV, Babu GS, Padmavathi M, Kumar BR, Alex RM, Schuhly W, Simic N, Kuhnelt D, Rao AV. Novel steroidal glycosides from two Indian *Caralluma* species, *C. stalagmifera* and *C. indica*.*Helvetica Chimica Acta*.2006; 89:201-209.
19. Al-Bekairi, AM, Qureshi S, Ahmed MM, Qazi NS, Khan ZA, Shah AH. Effect of *Caralluma tuberculata* on the cytological and biochemical changes induced by cyclophosphamide in mice. *Food Chem. Toxicol*.1992;30:719-722.
20. Abdel-Sattar E, Harraz FM, Al-Ansari SM, El-Mekkawy S, Ichino C, Kiyohara H, Ishiyama A, Ootoguro K, Omura S, Yamada H. Acylated pregnane glycosides from *Caralluma tuberculata* and their antiparasitic activity.*Phytochemistry*.2008;69:2180-2186.
21. Halaweish FT, Huntimer E, Khalil AT. Polyoxypregnane glycosides from *Caralluma retrospiciens*.*Phytochem Anal*.2004;15:189-194.
22. De Leo M, De Tommasi N, Sanogo R, Autore G, Marzocco S, Pizza C, Morelli I, Braca A. New pregnane glycosides from *Caralluma dalzielii*. *Steroids*.2005;70:573-585.
23. Bensuzan K. Taxonomy and conservation status of Moroccan stapeliads (Apocynaceae-Asclepiadoideae-Ceropegieae-Stapeliinae), *Bulletin de l'Institut Scientifique, Rabat.section Sciences de la Vie*.2009;31: 67-77.
24. Al-Khulaidi AA. *Flora of Yemen*, (SEMP, YEM/97/100)EPC, Sana'a, Yemen.2013.
25. Cassilas E, Ames W. Hepatotoxic effects of CCl<sub>4</sub> on English sole (*Parophrys vetulus*): Possible indicators of liver dysfunction. *Comp Biochem Physiol*.1986;84: 397-400.
26. Kotsanis N, Metcalfe CD. Enhancement of hepatocarcinogenesis in rainbow trout with carbon tetrachloride.*Bull Environ Contam Toxicol*.1991;46: 879-886.
27. Thrall KD, Vucelick ME, Gies RA, Zanger RC, Wietz KK, Poet TS, Springer DL, Grant DM, Benson GM. Comparative metabolism of carbon tetrachloride in rats, mice and hamsters using gas uptake and PBPK modeling.*J Toxicol Environ Health*.2000;60(8): 531-48.
28. Ogeturk M, Kus I, Colakoglu N, Zararsiz I, Ilhan N, Sarsilmaz M. Caffeic acid phenethyl ester protects kidneys against carbon tetrachloride in rats. *J Ethnopharmacol*.2005;97: 273-280.
29. Muriel P, Fernandez-Martinez E, Perez-Alvarez V, Lara-Ochoa F, Ponce S, Garcia J, Shibayama, M, Tsutsumi, V. Thalidomide ameliorates CCl<sub>4</sub>-cirrhosis in the rat.*Eur J Gastroenterol Hepatol*.2003;15: 951-957.
30. Moreno MG, Muriel P. Inducible nitric oxide synthase is not essential for the development of fibrosis and liver damage induced by CCl<sub>4</sub> in mice. *J Appl Toxicol* .2006;26: 326-332.
31. Venukumar MR, Latha MS. Hepatoprotective effect of the methanolic extract of orchids in Carbon tetrachloride treated male rats. *J Pharmacol*. 2002;34:2.
32. Bansa A, Adeyemo S. Phytochemical screening and antimalarial assessment of *Abutilon mauritianum*, *Bacopa monnifera* and *Datura stramonium*, *Biochemistry*.2006;18(1): 39 – 44.
33. Harborne JB. *Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis*, 2nd ed. Chapman and Hall, London.1984.
34. Trease GE, Evans WC. *A Textbook of Pharmacognosy*, 16th edn. Bailliere Tindall Ltd.: London.2009.
35. OECD, OECD Guideline 425: Acute Oral Toxicity—Up-and-Down Procedure. *OECD Guidelines for the Testing of Chemicals*, vol.2. Organization for Economic Cooperation and Development, Paris, France.2001.

36. Yang YS, Ahn TH, Lee JC, Moon CJ, Kim SH, Jun W, Park SC, Kim HC, Kim JC. Protective effects of Pycnogenol on carbon tetrachloride-induced hepatotoxicity in Sprague-Dawley rats. *Food Chem Toxicol.*2008; 46: 380–387.
37. Nema AK, Agarwa A, Kashaw V. Hepatoprotective activity of *Leptadenia reticulata* stems against carbon tetrachloride-induced hepatotoxicity in rats. *Indian J Pharmacol.*2011;43 (3):254-257.
38. Omar TY. Protective efficacy of *Glycyrrhiza glabra* on CCl<sub>4</sub>-induced liver injury in rabbits. *World J Pharm Res.*2014;3(3): 3627-3638.
39. Dhumal MS, Mane IH, Patel SS. Effect of Liv.52 on Carbon Tetrachloride-Induced Hepatotoxicity in Rabbits. *Indian J Pharmacol.*1989; 21:96.
40. Deliorman OD, Aslan M, Aktay G, Ergun E, Yesilada E, Ergun F. Evaluation of hepatoprotective effect of *Gentiana olivieri* herbs on subacute administration and isolation of active principle. *Life Sci.*2013;72: 2273-83.
41. Rage N, Dahanukar S, Karandikar SM. *Indian drugs.*1989. 27:18.
42. Dongare PP, Dhande SR, Kadam VJ. Standardization of Carbon Tetrachloride-Induced Hepatotoxicity In the Rat. *AJPTR.*2013; 3(5): 2249-3387.
43. Prochezian E, Ansari SH. Hepatoprotective activity of *Abutilon indicum* on experimental liver damage in rats. *Phytomedicine.*2005; 12: 62-64.
44. Ross MH, Reith EJ, Romrell LJ. *Histology: A Text and Atlas (2nd ed).* Baltimore, Williams & Wilkins.1989;51-84.
45. French SW, Miyamoto K, Ohta Y, Geoffrion Y. Pathogenesis of experimental alcoholic liver disease in the rat. *Methods Achiev Exp Pathol.*2000;13:181-207.
46. Pattanayak s, Nayak SS, Panda DP, Dinda SC, Shende V, Jadav. A hepatoprotective activity of crude flavonoid extract of *Cajanus Scarabaeoides*(L) in paracetamol intoxicated albino rats. *AJRBPS.* 2011;1(1):22-27.
47. Singh D, Gupta RS. Hepatoprotective activity of methanol extract of *Tecomella undulate* against alcohol and paracetamol induced hepatotoxicity in rats. *Life Sci Med Res.*2011;3:87-95.
48. Gould KS, Lister C. Flavonoid function in plants. In: Andersen OM, Markham KR, eds. *Flavonoids: Chemistry :biochemistry, and applications*, CRC press: Boca Raton.2006;397-441.
49. Hilaly JE, Israili ZH, Lyoussi B. Acute and chronic toxicological studies of *Ajugaivain* experimental animals. *J Ethnopharmacol.*2004;91: 43-50.
50. Mukinda JT, Eagles PF. Acute and sub-chronic oral toxicity profiles of the aqueous extract of *Polygala fruticosa* in female mice and rats. *J Ethnopharmacol.*2010;128: 236-240.
51. Kaplowitz N. Mechanisms of liver cell injury. *J Hepatol.*2000; 32: 39-47.
52. Bissel DM, Gores GJ, Laskin DL, Hoorhagle JH. Drug-induced liver injury: Mechanism and test systems. *Hepatol.*2001; 33: 1009-1013.
53. Pradeep HA. Hepatoprotective evaluation of *Anogeissus latifolia*: In vitro and in vivo studies. *World J Gastroentero.*2009;15(38):4816-22.
54. Wagner HK, Wolff PM. *New Natural Products and Plant Drugs with Pharmacological, Biological Or Therapeutical Activity: Proceedings of the First International Congress on Medicinal Plant Research, Section A, Held at the University of Munich, Germany, September 6-10, 1976.* Springer London, Limited.2011.
55. Fang YZ, Yang, S, Wu G. Free radicals, antioxidants, and nutrition. *Nutrition.*2002;18(10):872–9.
56. Basu S. Carbon tetrachloride-induced lipid peroxidation:Eicosanoid formation and their regulation by antioxidant nutrients, *Toxicology.*2003; 189:113-27.
57. Manibusan MK, Odin M, Eastmond DA. Postulated carbon tetrachloride mode of action: a review. *J Environ Sci Health C Environ Carcinog Ecotoxicol Rev.*2007;25:185-209.
58. Planaguma A, Claria J, Miquel R, Lopez-Parra M, Titos E, Masferrer JL et al. The selective cyclooxygenase-2 inhibitor SC-236 reduces liver fibrosis by mechanisms involving non-parenchymal cell apoptosis and PPAR gamma activation, *FASEB.*2005;19:1120-2.
59. Sureshkumar SV, Mishra SH. Hepatoprotective effect of extracts from *Pergula riadaemia* Forsk. *J Ethnopharmacol.*2006;107: 164-168.
60. Kozer E, Evans S, Barr J, Greenberg R, Soriano I, Bulkowstein M, Petrov I, Chen-Levi Z, Barzilay B, Berkovitch M. Glutathione, glutathione-dependent enzymes and antioxidant status in erythrocytes from children treated with high-dose paracetamol. *Br J Clin Pharmacol.*2003;55(3), 234- 40.
61. Girish C, Koner BC, Jayanthi S, Rao KR, Rajesh B, Pradhan SC. Hepatoprotective activity of six poly herbal formulations in paracetamol-induced liver toxicity in mice. *Indian J Med Res.*2009;129:569-578.
62. Brent JA, Rumack BH. Role of free radicals in toxic hepatic injury II. *Clin Toxicol.*1993;31: 173–196.

63. Zhang JS, Wang HL, Yan XX, Zhang LD. Comparison of short-term toxicity between nano-Se and selenite in mice. *Life Sci.* 2005;76:1099-109.
64. Rajesh MG, Latha MS. Preliminary evaluations of the antihepatotoxic effect of Kamilari, apolyherbal formulation. *J Ethnopharmacol.* 2004;91:99-104.
65. Naik SR, Panda VS. Hepatoprotective effect of Ginkgo select phytosome in rifampicin induced liver injury in rats: evidence of antioxidant activity. *Fitoterapia* .2008;79:439-45.
66. Al-Mehdar AA, El-Denshary ES, Abdel-Wahhab MA. Alpha lipoic acid and alpha-tocopherol counteract the oxidative stress and liver damage in rats sub-chronically treated with khat (*Catha edulis*) Extract. *Global J. of Pharmacology.* 2012;6(2):94-105.
67. Mukherjee PK. Plant products with hypercholesterolemic potentials', In: Taylor, Steve L. (Ed.), *Advance in Food and Nutrition Research*, 47. Elsevier Science, USA. 2003;277-338.
68. Ranawat L, Bhatt J, Patel J. Hepatoprotective activity of ethanolic extracts of bark of *Zanthoxylum armatum* DC in CCl<sub>4</sub> induced hepatic damage in rats. *J Ethnopharmacol.* 2010;127:777-780.
69. Aniya Y, Koyama T, Miyagi C. Free radical scavenging and hepatoprotective actions of the medicinal herb, *Crossocephalum crepidioides* from the Okinawa Islands. *Biol Pharm Bull.* 2005;28(1):19-23.
70. Suresh Kumar SV, Sujatha C, Syamala J, Nagasudha B, Mishra SH. Hepatoprotective activity of extracts from *Pergularia adaemia Forsk* against Carbon tetrachloride induced toxicity in rats. *Phcog Mag.* 2007;3:11.
71. Bishayee A, Sarkar A, Chatterjee M. Hepatoprotective activity of carrot (*Daucus carota* L.) against carbon tetrachloride intoxication in mouse liver. *J Ethnopharmacol.* 1995;47:69-74.
72. Soni B, Visavadiya N, Madamwar D. Ameliorative action of *cyanobacterial phycoerythrin* on CCl<sub>4</sub>-induced toxicity in rats. *Toxicol.* 2008;248: 59-65.
73. Gowda S, Desai PB, Kulkarni SS, Hull VV, Math AA, Vernekar SN. Markers of renal function tests. *J Med Sci.* 2010;2(4):170-173.
74. Rincón AR, Covarrubias A, Pedraza-Chaverri J, Poo JL, Armendáriz-Borunda J, Panduro A. Differential effect of CCl<sub>4</sub> on renal function in cirrhotic and non-cirrhotic rats. *Exp Toxicol Pathol.* 1999;51(3): 199-205.
75. Rahmat AA, Dar FA, Choudhary IM. Protection of CCl<sub>4</sub>-Induced Liver and Kidney Damage by Phenolic Compounds in Leaf Extracts of *Cnestis ferruginea* (de Candolle). *Pharmacognosy Res.* 2014;6(1):19-28. doi:10.4103/0974-8490.122913.
76. Preethi KC, Kuttan R. Hepato and reno protective action of *Calendula Officinalis* L. flower extract. *Indian J Exp Biol.* 2009;47:163-168.
77. Al-Duais AM, Al-Awthan YS, Mukhtar AA, Shamsan AA. Prevention of Carbon Tetrachloride (CCl<sub>4</sub>)-Induced Liver Damage in Guinea Pigs by *Cyphostemma Digitatum*. *J Life Scis.* 2012;6:137-43.
78. Mion F, Geloën A, Agosto E, Minaire Y. Carbon tetrachloride-induced cirrhosis in rats influence of the acute effects of the toxin on glucose metabolism. *Hepatol.* 1996;23(3):582-588.
79. Muriel P, Alba N, Perez-Alvarez VM, Shibayama M, Tsutsumi VM. Kupffer cells inhibition prevents hepatic lipid peroxidation and damage induced by carbon tetrachloride. *Comp Biochem Physiol C Toxicol Pharmacol.* 2001;130(2): 219-226.
80. Gaw A, Cowan RA, O'Reill DS, Stewart MJ, Shepherd J. *Clinical Biochemistry-An Illustrated Color Text*, Churchill Livingstone, Oxford. 1998;56
81. Sayantan R, Abhishek S. Antidiabetic activity of *Caralluma edulis* bark and leaf extract against streptozotocin induced diabetic rats. *NSHMJ Pharm Healthcare Management.* 2012;03:76-81.
82. Saini MR, Kumar S, Jagetia GC, Navita S. Effectiveness of Liv-52 against radiation sickness and dermatitis. *Indian Pract.* 1984;1133.
83. Junnila M, Rahko T, Sukura A. Reduction of carbon tetrachloride-induced hepatotoxic effects by oral administration of betaine in male Han-wistar rats: a morphometric histological study. *Vet Pathol.* 2003;37:231-238.
84. Karakus E, Karadeniz A, Simsek N. Protective effect of Panax ginseng against serum biochemical changes and apoptosis in liver of rats treated with carbon tetrachloride (CCl<sub>4</sub>). *J Hazard Mater.* 2011; 195:208-213.